

**Genetic Stock Identification of Chinook
Salmon caught in Northern British Columbia
Troll Fisheries 2017**

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ABSTRACT

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Fisheries & Oceans Canada has managed the Northern British Columbia (NBC) troll fishery since 1995 to reduce impacts on Chinook salmon (*Oncorhynchus tshawytscha*) stocks from the West Coast of Vancouver Island (WCVI). Microsatellite DNA based stock identification techniques have been used to address stock specific management in this mixed stock fishery since 2002. Chinook salmon stock compositions were estimated for the 2017 NBC troll fishery and the fishery was managed in-season to a harvest rate target for WCVI Chinook salmon. The application of stock specific management allowed the internationally negotiated catch allocation to be attempted while reducing the exploitation of WCVI Chinook. Stock compositions were applied to catch to provide region specific estimates of the impact of the 2017 NBC troll fisheries on all of the Chinook stock groups encountered.

INTRODUCTION

Funding for this project was provided by the Pacific Salmon Commission's Northern Boundary and Transboundary Rivers Restoration and Enhancement Fund (Northern Fund) to estimate Chinook salmon (*Oncorhynchus tshawytscha*) stock compositions in Northern British Columbia (NBC) troll fisheries in 2017. This report presents estimates of the Chinook salmon stock identification based on genetic analyses of samples collected from the fishery. The results are part of a continuing project to examine the genetics of Chinook salmon caught in NBC Troll fisheries. This document fulfills the reporting requirements for the 2017 Northern Fund project. Costs to the Northern Fund consisted of the genetic analyses and a portion of the sampling costs. Other components of the project were funded by Fisheries & Oceans Canada in existing programs.

The primary objective of this study was to estimate stock specific catches by the NBC Troll fishery in 2017. The commercial troll fishery is the largest fishery for Chinook salmon in NBC and is one of two fisheries defined within the Aggregate Abundance Based Management (AABM) regime implemented by the Pacific Salmon Treaty (PST)(2000) for the North Coast. Under the revised international agreement the NBC troll and Haida Gwaii (QCI) sport fisheries are managed in aggregate within the same regime. Current domestic allocation policies within Canada require the NBC troll fishery to be influenced first by any management actions required to protect weak stocks (Winther and Beacham 2006).

The North Coast troll fishery has been defined as Area F consisting of Pacific Fishery Management Areas 1 to 10, 101 to 110, and 130 and 142 (Figure 1.). The AABM portion of the NBC troll fishery makes up the north-western portion of Area F; Areas 1 to 5, 101 to 105, 130 and 142. Historically the size of the troll fleet in Area F was relatively stable from 2002 to 2005, varying between 146 and 168 licensed vessels. Through a series of area selection processes the Area F fleet increased to 284 licensed vessels from 2008 to 2010 and then declined to 238 in 2015. The change in fleet size was initially due to reduced fishing opportunities in southern areas and the introduction of individual transferable quotas (ITQ) in the NBC troll fishery for Chinook salmon in 2005. The number of vessels fishing is considerably less than the licenses available in Area F, for example 143 vessels fished in 2017.

The application of DNA-level markers for stock identification in the NBC troll fishery has continued since 2002. After moderate success in the initial year the experiments essentially met their objective of allowing the Chinook salmon quota negotiated under the PST agreement to be harvested while continuing to reduce impacts on stocks of conservation concern from the West Coast of Vancouver Island (WCVI) (Winther and Beacham 2006 & 2009). Subsequent fisheries were less successful averaging only 72% of the pre-fishery allowance to the troll fishery. These lower catches were the result of forecasting errors and policy changes. A series of under forecasts of WCVI Chinook lead to early closures of the NBC Troll fishery after 2006. The movement of troll licenses to the Pacific Integrated Commercial Fisheries Initiative (PICFI), the Allocation Transfer Program (ATP) and other processes essentially removed licenses from fishing in Area F while leaving their allocation within the ITQ. Consequently a portion of the quota was not available to be fished and Area F could not reach the allowable catch determined under the Treaty.

The 2017 pre-season Aggregate Abundance Based Management (AABM) index was 1.15 with an associated allowable catch of 149,500 Chinook salmon for NBC Troll and QCI sport fisheries. The pre-season estimate of the sport catch was 41,000 fish leaving 108,500 fish as the pre-season troll catch target. The in-season target for the NBC troll fishery was a maximum harvest rate of 3.2% on Chinook salmon of WCVI origin returning to Canadian waters.

A demonstration fishery to examine the application of individual transferable quotas (ITQ) in the troll fishery was first held in 2005 and was continued in 2017. The pre-season troll allocation was divided among 239 licenses for an individual quota of 454 Chinook salmon per licence at the beginning of the season.

Stock identification is a key component in the management of mixed stock salmon fisheries. The application of DNA-level markers for stock identification, particularly microsatellites, has provided much greater resolution among Chinook salmon populations than was possible with previous genetic markers (Beacham et al. 1996; Banks et al. 2000; Beacham et al. 2003). If the baseline used to estimate stock composition is adequate, microsatellites can be applied successfully on a local basis to provide information on stock composition even when there is a complex mixture of populations in the catch (Beacham et al. 2006).

We used microsatellite variation in Chinook salmon as a tool for management of stocks of conservation concern encountered by the NBC troll fishery. Our challenge was to provide advice to managers that would allow the Northern British Columbia (Area F) troll fishery to maximize catch of Chinook salmon while minimizing the exploitation of WCVI Chinook salmon. The biological sampling objectives were to generate stock compositions for the troll catch of Chinook salmon in NBC.

METHODS

Stock specific catch estimates were generated from the results of genetic samples applied to catch data. Genetic samples were grouped temporally and weighted to the catch for the periods sampled. In cases where the temporal strata for samples overlapped, the daily catch during the overlapping days were weighted to each sample based on the relative number of fish in each sample.

Two programs provided catch accounting of Chinook salmon from the NBC Troll fishery; daily catch records (hails) and landing validation. Vessel operators must provide a record of their daily catch either electronically or by phone to the Fisheries Operating System as a condition of licence. Electronic or phone-in records were required within 24 hours of landing or within 24 hours of the closure of the fishery and consisted of date fished, area fished, number of fish caught and retained by species and number of fish caught and released by species. In addition, all vessel landings of Chinook salmon must be counted and reported (validated) by the third party contractor. The combination of daily records reported by the fishers and validation by the third party provide records that are essentially a census of the Chinook salmon catch. Catch records were treated as a census without variance.

The sample design was approached from the perspective of a binomial problem where Chinook salmon were identified as either from WCVI or not. If the WCVI component was near 5% and the catch approached the ceiling of 108,500 then the level of precision afforded by ~2,600 samples would provide 95% confidence limits plus or minus ~20% of the estimated proportion of WCVI Chinook. Smaller proportions would have respectively broader confidence limits and smaller sample sizes would have respectively broader confidence limits as well.

The program to sample commercial fishery landings was designed with the objective of collecting tissues from approximately 2.7% (~3000 fish) of the Chinook salmon caught in the fishery. Samples were collected by a contractor involved in the existing MRP sampling program. The collection target was designed larger than the target for analysis (2,600 fish) because sample collection was relatively cheap when compared with genetic analyses. The extra samples also allowed for sub-sampling such that the samples submitted for analyses would best represent the commercial fishery landings across the time and area strata in the fishery.

Ultimately 2,471 fish were sampled, 2,089 fish were submitted for analyses and results were received from 2,071 samples.

The NBC Troll fishery was sampled by a third party contractor. The contract description included random selection of troll vessel landings with stratified sampling of fish within the selected landing. The sampling procedure was to select vessel landings at random and sample less than 50 fish from each delivery. Every 5th or 10th fish was sampled from the load depending on the size of the delivery to spread the samples through the load and to reduce autocorrelation. Individual Chinook salmon sampled from commercial fishery landings were sampled for nose-fork length and scales. Fish landed by the commercial fishery were dressed so gender could not be determined. These collections were matched to data on the area fished and date caught. An identification number was included for fish sampled that had coded wire tags (Appendix 1).

Scale samples were collected on to scale books, five scales per fish, as described by MacLellan (1999). Data on the geographic location, date, and sampler accompanied each sample. Samples were forwarded to the Fisheries & Oceans Canada, Molecular Genetics Laboratory at the Pacific Biological Station in Nanaimo.

Chinook salmon collections were compared against genetic baselines from 302 Chinook salmon populations from Southeast Alaska through Canada and the lower United States of America (Appendix 2). Samples were analyzed for 15 microsatellite loci using methods of DNA extraction, PCR reaction, electrophoresis, and allele scoring described by Candy et al. (2002) and Beacham et al. (2006).

The Molecular Genetics Laboratory provided the sample analysis. A new version of the computer program as outlined by Pella and Masuda (2001) was developed and used for the analyses presented here. The program CBAYES (Neaves et al 2005) can be downloaded from the Molecular Genetics Laboratory website (Fisheries & Oceans Canada, Molecular Genetics Laboratory, Pacific Biological Station, Nanaimo). The model output presented includes the Bayesian probability estimates for the 5 most probable populations for each sample.

RESULTS

The NBC troll fishery landed 97,730 Chinook salmon in 2017. Chinook salmon fishing was permitted from June 21 to August 4 and from August 25 to September 30. Chinook salmon catches included 88,991 fish in the June to August fishery and 8,739 fish in the August and September fishery. Daily catch was highest on opening day, June 21, when 103 active vessels caught 6,880 Chinook salmon. Catches declined quickly and were below 2,500 fish per day after the first 3 days of the fishery (Figure 2.).

Genetic samples were collected from 172 of the 473 vessel landings that unloaded Chinook salmon from the 2017 Troll fishery. The collection was sub-sampled and 2,089 fish samples from 140 vessel landings were submitted for analyses. The samples were submitted and reported in 6 groups. The first 5 groups were from the June to August fishery and represented catches from periods ranging from 8 to 14 days. Since fishing was continuous from June 21 to August 4, the groups of samples overlapped for periods up to 8 days. The samples from the September fishery were submitted as a single group representing 37 days of fishing. In September catch declined to near zero at the end of the month.

The most prevalent Chinook salmon stock group in the NBC troll catch was the South Thompson (SOTH) stock group making up 31.6% of the catch or 30,868 fish. Other significant components were Upper Columbia Summer & Fall (Up Col-Su/F), North & Central Oregon, and Coastal Washington stock groups (Table 1). This pattern persisted through the samples collected from the June to August fishery but changed in September as the SOTH fish declined significantly and the Columbia and Oregon components increased (Table 2 and Table 3.).

Chinook salmon from WCVI were most prevalent in the August 25 to September 30 samples at 11.8%. They were least prevalent in the June 26th to July 8th sample making up 2.5% of the sample. A total of 4,578 Chinook salmon from WCVI were estimated to be caught by the fishery.

DISCUSSION

The NBC troll fishery landed 97,730 Chinook salmon in 2017. While there were potentially 108,500 fish available to the troll fishery (149,500 total AABM allowable catch less 41,000 QCI sport catch), catches were limited by the ITQ program. The quota associated with approximately 21 licenses held by PICFI and ATP programs was not fished (~9,500 Chinook). The fishery was managed for concerns for WCVI Chinook salmon but the only management action in this regard was to close the fishery from August 5 to 24.

Catch per unit effort (CPUE) is difficult to interpret for the Chinook troll fishery because of the influence of ITQ's on fishing behavior. CPUE's was high on opening day but declined quickly. CPUE on opening day, June 21, was 67 Chinook salmon per boat day. CPUE dropped to 45 on the second day and then declined to approximately 24 fish per boat day by the end of the week. CPUE increased at the end of June and remained between 30 and 40 fish per boat day for the first week of July.

Recreational fisheries have priority access to Chinook salmon so the shares within the AABM allowable catch are determined pre-season by forecasting the recreational catch. The forecast for the AABM recreational catch of 41,000 underestimated the amount of fish caught. The preliminary sport catch estimate for 2017 was 45,600 Chinook salmon. The recreational catch brings the preliminary estimate for the total AABM catch to 143,330.

Canada's domestic management target for the NBC Troll fishery is to have an exploitation rate (ER) of less than 3.2% on WCVI Chinook salmon. Since Alaskan harvests are unknown in-season this ER is approximated by a harvest rate of 3.2% on the return of WCVI Chinook salmon to Canadian waters. The estimate of WCVI Chinook salmon caught by the troll fishery using genetic samples was 4,578 fish. The actual harvest rate on WCVI Chinook will not be known until the post-season fishery and escapement data have been analyzed.

Summer troll fisheries have been excluded from areas around Langara Island and the north end of Graham Island since 1997 to avoid conflicts with the sport fishery. The ribbon boundary that keeps the troll fishery away from the shore has assisted in the objective of avoiding WCVI Chinook stocks since those fish tend to migrate closer to the shore (Winther and Beacham, 2006 & 2009).

The combination of Chinook salmon abundance and high proportions of WCVI fish makes troll fishery impacts on WCVI stocks most severe in August (Winther and Beacham, 2006 & 2009). The second portion of the 2017 troll fishery was opened August 25 as in 2015 and 2016. A sample collected from catches between August 25 and September 30 also had the highest component of WCVI Chinook salmon of the season at 11.8% (Table 2.). The majority of the fish caught in the second opening were caught during the beginning of the opening. Catches during the first 14 days of the fishery accounted for 8,133 of the 8,739 fish caught over the entire August 25 to September 30 fishery.

The timing differential between WCVI Chinook salmon and abundant stocks in the Fraser River and Upper Columbia provided opportunities to minimize impacts on WCVI stocks by fishing in June and July. The increase in Columbia Summer/Fall stocks doesn't appear to have changed these conditions of higher WCVI proportions in August. The sample collected from catches in late August and September had the highest amount of WCVI Chinook in the

stock mixtures (11.8%). Typically samples from late September have few fish from WCVI but the fishing opportunities in September are hampered by bad weather and the decline of Chinook abundance through the month.

Differences in timing and distribution between stock groups have allowed the fishery to be positioned such the stocks of conservation concern were avoided and more abundant stocks were harvested. Changes in abundance or distribution of either the stock of conservation concern or any of the major components encountered by the fishery could have significant consequences. The ability to have a successful mixed stock fishery while avoiding a stock of conservation concern hinges on identifying such differences.

The application of DNA level markers for stock identification to the management of a particular stock within a mixed stock fishery depends heavily on the ability to identify the stock and detect it with a reasonable level of precision. The management targets and proportions of WCVI Chinook salmon experienced in 2017 are near the edge of the effective range of the technique given the sample sizes that the program could afford (i.e. proportions near 5%). In addition, the relative proportions of component stocks present in the fishery must show some variability across time or space. If the proportion of a stock of conservation concern was static or random across the fishery, fishing periods or areas could not be defined that were any better for avoiding the stock. This could happen if the stock of conservation concern had the same or similar timing and distribution as abundant stocks within the fishery.

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TABLES

Table 1. Chinook salmon catch by stock group for 2017 NBC troll fisheries.

Standard deviations (STD) appear in brackets.

Code	Region	Proportion (%)	STD (%)	Catch	STD
1	Upper Fraser (UPFR)	0.2	(0.1)	149	(118)
2	Middle-Upper Fraser (MUFR)	0.8	(0.3)	756	(257)
3	Lower Fraser Fall (LWFR-F)	0.1	(0.1)	138	(79)
4	North Thompson (NOTH)	0.4	(0.2)	430	(171)
5	South Thompson (SOTH)	31.6	(1.1)	30,868	(1059)
6	Lower Thompson (LWTH)	0.0	(0.0)	0	(22)
7	East Coast Vancouver Island (ECVI)	1.4	(0.4)	1,371	(358)
8	West Coast Vancouver Island (WCVI)	4.7	(0.5)	4,569	(466)
9	Southern BC Mainland (SOMN)	0.6	(0.2)	589	(192)
10	Northern BC Mainland (NOMN)	1.6	(0.4)	1,555	(343)
11	Nass	0.1	(0.1)	49	(58)
12	Lower Fraser Spring (LWFR-Sp)	0.2	(0.1)	209	(111)
13	Lower Fraser Summer (LWFR-Su)	0.0	(0.0)	40	(41)
14	Haida Gwaii (QCI)	0.0	(0.1)	48	(50)
15	Alaska	0.0	(0.1)	39	(78)
17	Taku	0.1	(0.1)	49	(85)
18	Stikine	0.1	(0.1)	68	(99)
19-23	Skeena	1.2	(0.3)	1,156	(306)
24	Alsek	0.0	(0.0)	31	(39)
50	Puget Sound	1.4	(0.3)	1,383	(295)
51	Juan de Fuca	0.3	(0.1)	256	(140)
52	Coastal Washington	8.2	(0.7)	7,984	(675)
53	Lower Columbia	2.2	(0.4)	2,167	(390)
54	Upper Columbia Spring	0.0	(0.0)	0	(16)
55	Upper Columbia Summer/Fall	20.6	(1.1)	20,173	(1045)
56	Snake Spring/Summer	0.0	(0.0)	18	(40)
57	Snake Fall	4.3	(0.7)	4,198	(680)
58	North & Central Oregon	14.6	(0.9)	14,261	(907)
59	South Oregon Coastal	3.3	(0.6)	3,205	(576)
61	Klamath/Trinity	0.0	(0.0)	5	(28)
62	Middle Columbia Spring	0.1	(0.1)	58	(98)
63	Upper Willamette	1.8	(0.4)	1,789	(396)
64	Central Valley Fall	0.1	(0.1)	60	(65)
65	Central Valley Spring	0.1	(0.1)	57	(79)
66	Coastal California	0.0	(0.0)	0	(8)
	TOTAL	100%		97,730	

Table 2. Chinook stock proportions observed in samples from 2017 NBC Troll catches.

Composition presented as % of the sample N. Standard deviations appear in brackets.

	Year	2017		2017		2017		2017		2017	
	Date	June 21-July 3		June 16-July 8		July 7-July 14		July 12-July 20		July 22-Aug 4	
	Catch	29,398		25,913		11,230		12,052		10,398	
Code	N	371		544		302		298		247	
		%	STD	%	STD	%	STD	%	STD	%	STD
1	Upper Fraser (UPFR)	0.1	(0.2)	0.2	(0.3)	0.2	(0.3)	0.3	(0.3)	0.2	(0.4)
2	Middle-Upper Fraser (MUFR)	1.3	(0.7)	1.0	(0.6)	0.2	(0.3)	0.4	(0.4)	0.4	(0.5)
3	Lower Fraser Fall (LWFR-F)	0.0	(0.0)	0.0	(0.0)	0.0	(0.0)	0.0	(0.0)	0.6	(0.6)
4	North Thompson (NOTH)	0.1	(0.3)	0.6	(0.4)	1.2	(0.7)	0.3	(0.4)	0.6	(0.6)
5	South Thompson (SOTH)	28.5	(2.4)	40.2	(2.2)	36.5	(2.8)	33.5	(2.8)	30.8	(3.0)
6	Lower Thompson (LWTH)	0.0	(0.0)	0.0	(0.0)	0.0	(0.1)	0.0	(0.1)	0.0	(0.1)
7	East Coast Vancouver Island (ECVI)	2.1	(1.0)	1.0	(0.4)	1.0	(0.6)	2.1	(0.9)	0.9	(0.6)
8	West Coast Vancouver Island (WCVI)	4.1	(1.0)	2.5	(0.7)	2.6	(0.9)	4.7	(1.2)	7.8	(1.7)
9	Southern BC Mainland (SOMN)	0.5	(0.4)	0.6	(0.4)	2.1	(0.9)	0.5	(0.5)	0.0	(0.2)
10	Northern BC Mainland (NOMN)	2.0	(0.8)	2.4	(0.8)	0.7	(0.6)	1.4	(0.7)	1.0	(0.7)
11	Nass	0.0	(0.1)	0.0	(0.1)	0.0	(0.2)	0.4	(0.4)	0.0	(0.1)
12	Lower Fraser Spring (LWFR-Sp)	0.3	(0.3)	0.2	(0.2)	0.3	(0.3)	0.0	(0.1)	0.4	(0.4)
13	Lower Fraser Summer (LWFR-Su)	0.0	(0.0)	0.0	(0.0)	0.0	(0.0)	0.3	(0.3)	0.0	(0.0)
14	Haida Gwaii (QCI)	0.0	(0.0)	0.2	(0.2)	0.0	(0.0)	0.0	(0.0)	0.0	(0.0)
15	Alaska	0.0	(0.1)	0.1	(0.3)	0.0	(0.1)	0.0	(0.1)	0.0	(0.1)
17	Taku	0.0	(0.2)	0.0	(0.2)	0.0	(0.1)	0.1	(0.2)	0.1	(0.4)
18	Stikine	0.0	(0.1)	0.1	(0.3)	0.1	(0.2)	0.2	(0.4)	0.0	(0.1)
19-23	Skeena	1.1	(0.7)	2.0	(0.7)	0.7	(0.8)	2.0	(0.9)	0.0	(0.2)
24	Alsek	0.0	(0.1)	0.0	(0.0)	0.0	(0.0)	0.0	(0.1)	0.0	(0.1)
50	Puget Sound	0.3	(0.4)	1.6	(0.6)	1.1	(0.7)	2.3	(1.0)	2.6	(1.3)
51	Juan de Fuca	0.8	(0.5)	0.0	(0.0)	0.0	(0.0)	0.1	(0.3)	0.0	(0.0)
52	Coastal Washington	7.0	(1.5)	7.4	(1.2)	8.6	(1.8)	8.8	(1.8)	11.5	(2.2)
53	Lower Columbia	2.7	(0.9)	2.3	(0.8)	0.2	(0.5)	2.6	(1.0)	1.7	(0.9)
54	Upper Columbia Spring	0.0	(0.0)	0.0	(0.0)	0.0	(0.0)	0.0	(0.0)	0.0	(0.0)
55	Upper Columbia Summer/Fall	21.5	(2.4)	15.3	(1.8)	22.6	(2.7)	18.2	(2.6)	20.3	(3.0)
56	Snake Spring/Summer	0.0	(0.1)	0.0	(0.1)	0.0	(0.1)	0.0	(0.1)	0.0	(0.1)
57	Snake Fall	5.8	(1.7)	4.1	(1.2)	0.8	(1.3)	2.4	(1.5)	5.5	(2.0)
58	North & Central Oregon	15.6	(2.1)	12.6	(1.6)	17.7	(2.5)	12.0	(2.4)	9.7	(2.1)
59	South Oregon Coastal	2.6	(1.3)	2.5	(0.9)	3.1	(1.3)	7.3	(2.1)	5.7	(1.8)
61	Klamath/Trinity	0.0	(0.0)	0.0	(0.1)	0.0	(0.0)	0.0	(0.0)	0.0	(0.1)
62	Middle Columbia Spring	0.1	(0.3)	0.0	(0.0)	0.3	(0.4)	0.0	(0.1)	0.0	(0.1)
63	Upper Willamette	3.4	(1.1)	2.9	(0.8)	0.0	(0.1)	0.1	(0.3)	0.0	(0.1)
64	Central Valley Fall	0.0	(0.1)	0.2	(0.2)	0.0	(0.1)	0.0	(0.1)	0.0	(0.1)
65	Central Valley Spring	0.2	(0.3)	0.0	(0.0)	0.0	(0.0)	0.0	(0.0)	0.0	(0.0)
66	Coastal California	0.0	(0.0)	0.0	(0.0)	0.0	(0.0)	0.0	(0.0)	0.0	(0.0)

Table 2 continued. Chinook stock proportions observed in samples from 2017 NBC Troll catches.

Composition presented as % of the sample N. Standard deviations appear in brackets.

	Year	2017	
	Date	Aug 25-Sep 30	
	Catch	8,739	
Code	N	309	
		%	STD
1	Upper Fraser (UPFR)	0.0	(0.1)
2	Middle-Upper Fraser (MUFR)	0.0	(0.1)
3	Lower Fraser Fall (LWFR-F)	0.8	(0.5)
4	North Thompson (NOTH)	0.0	(0.1)
5	South Thompson (SOTH)	8.2	(1.6)
6	Lower Thompson (LWTH)	0.0	(0.1)
7	East Coast Vancouver Island (ECVI)	0.5	(0.5)
8	West Coast Vancouver Island (WCVI)	11.8	(1.8)
9	Southern BC Mainland (SOMN)	0.0	(0.1)
10	Northern BC Mainland (NOMN)	0.1	(0.2)
11	Nass	0.0	(0.1)
12	Lower Fraser Spring (LWFR-Sp)	0.0	(0.0)
13	Lower Fraser Summer (LWFR-Su)	0.0	(0.0)
14	Haida Gwaii (QCI)	0.0	(0.0)
15	Alaska	0.0	(0.1)
17	Taku	0.1	(0.3)
18	Stikine	0.1	(0.3)
19-23	Skeena	0.0	(0.1)
24	Alsek	0.3	(0.3)
50	Puget Sound	2.5	(1.0)
51	Juan de Fuca	0.0	(0.0)
52	Coastal Washington	8.9	(1.8)
53	Lower Columbia	3.1	(1.1)
54	Upper Columbia Spring	0.0	(0.0)
55	Upper Columbia Summer/Fall	34.8	(3.1)
56	Snake Spring/Summer	0.2	(0.3)
57	Snake Fall	5.6	(1.9)
58	North & Central Oregon	22.4	(2.6)
59	South Oregon Coastal	0.1	(0.3)
61	Klamath/Trinity	0.0	(0.1)
62	Middle Columbia Spring	0.0	(0.1)
63	Upper Willamette	0.4	(0.4)
64	Central Valley Fall	0.0	(0.1)
65	Central Valley Spring	0.0	(0.0)
66	Coastal California	0.0	(0.0)

Table 3. Chinook catch composition from 2017 NBC troll fisheries by sample group.

Catch composition presented in numbers of fish. Standard deviations appear in brackets.

	Year	2017		2017		2017		2017		2017	
	Date	June 21-July 3		June 16-July 8		July 7-July 14		July 12-July 20		July 22-Aug 4	
	Catch	29,398		25,913		11,230		12,052		10,398	
Code	N	371		544		302		298		247	
		%	STD	%	STD	%	STD	%	STD	%	STD
1	Upper Fraser (UPFR)	19	(60)	51	(73)	18	(38)	41	(42)	21	(43)
2	Middle-Upper Fraser (MUFR)	386	(191)	265	(152)	17	(35)	46	(47)	41	(54)
3	Lower Fraser Fall (LWFR-F)	0	(10)	0	(7)	0	(5)	0	(5)	66	(62)
4	North Thompson (NOTH)	30	(87)	157	(95)	139	(81)	41	(43)	63	(66)
5	South Thompson (SOTH)	8382	(692)	10425	(559)	4097	(316)	4039	(336)	3208	(314)
6	Lower Thompson (LWTH)	0	(14)	0	(10)	0	(10)	0	(7)	0	(7)
7	East Coast Vancouver Island (ECVI)	606	(305)	264	(116)	110	(65)	255	(107)	90	(65)
8	West Coast Vancouver Island (WCVI)	1216	(308)	644	(179)	291	(103)	571	(149)	814	(177)
9	Southern BC Mainland (SOMN)	133	(121)	155	(92)	240	(101)	59	(57)	2	(16)
10	Northern BC Mainland (NOMN)	588	(230)	609	(212)	76	(71)	170	(90)	105	(76)
11	Nass	0	(21)	0	(14)	3	(20)	45	(47)	0	(9)
12	Lower Fraser Spring (LWFR-Sp)	82	(82)	48	(50)	37	(37)	0	(10)	42	(42)
13	Lower Fraser Summer (LWFR-Su)	0	(4)	0	(4)	0	(2)	40	(41)	0	(3)
14	Haida Gwaii (QCI)	0	(7)	48	(49)	0	(4)	0	(3)	0	(3)
15	Alaska	3	(27)	36	(72)	0	(6)	0	(7)	0	(6)
17	Taku	10	(49)	8	(39)	0	(8)	7	(29)	14	(40)
18	Stikine	1	(21)	28	(76)	6	(22)	24	(51)	1	(11)
19-23	Skeena	315	(206)	527	(179)	74	(86)	237	(107)	3	(21)
24	Alsek	3	(23)	0	(8)	0	(5)	0	(6)	0	(7)
50	Puget Sound	80	(132)	406	(148)	126	(80)	275	(125)	275	(135)
51	Juan de Fuca	238	(136)	0	(4)	0	(2)	18	(33)	0	(3)
52	Coastal Washington	2055	(433)	1929	(320)	970	(203)	1058	(217)	1195	(227)
53	Lower Columbia	794	(277)	586	(201)	26	(51)	308	(118)	180	(97)
54	Upper Columbia Spring	0	(11)	0	(7)	0	(5)	0	(5)	0	(5)
55	Upper Columbia Summer/Fall	6334	(711)	3956	(478)	2541	(302)	2193	(314)	2110	(308)
56	Snake Spring/Summer	0	(20)	0	(14)	0	(9)	0	(9)	0	(10)
57	Snake Fall	1696	(486)	1054	(318)	94	(141)	290	(185)	575	(209)
58	North & Central Oregon	4595	(627)	3273	(408)	1983	(279)	1445	(294)	1005	(218)
59	South Oregon Coastal	751	(395)	637	(231)	346	(148)	877	(257)	588	(185)
61	Klamath/Trinity	0	(11)	4	(23)	0	(4)	0	(4)	0	(6)
62	Middle Columbia Spring	24	(87)	0	(13)	34	(43)	0	(7)	0	(6)
63	Upper Willamette	1000	(325)	745	(219)	0	(6)	10	(35)	0	(6)
64	Central Valley Fall	1	(23)	58	(59)	0	(7)	0	(8)	0	(8)
65	Central Valley Spring	57	(78)	0	(7)	0	(5)	0	(5)	0	(5)
66	Coastal California	0	(4)	0	(4)	0	(2)	0	(2)	0	(3)

Table 3 continued. Chinook catch composition from 2017 NBC troll fisheries by sample group.

Catch composition presented in numbers of fish. Standard deviations appear in brackets.

	Year	2017	
	Date	Aug 25-Sep 30	
	Catch	8,739	
Code	N	309	
		%	STD
1	Upper Fraser (UPFR)	0	(8)
2	Middle-Upper Fraser (MUFR)	2	(13)
3	Lower Fraser Fall (LWFR-F)	72	(48)
4	North Thompson (NOTH)	0	(5)
5	South Thompson (SOTH)	718	(141)
6	Lower Thompson (LWTH)	0	(5)
7	East Coast Vancouver Island (ECVI)	46	(41)
8	West Coast Vancouver Island (WCVI)	1032	(161)
9	Southern BC Mainland (SOMN)	0	(6)
10	Northern BC Mainland (NOMN)	6	(20)
11	Nass	0	(6)
12	Lower Fraser Spring (LWFR-Sp)	0	(4)
13	Lower Fraser Summer (LWFR-Su)	0	(2)
14	Haida Gwaii (QCI)	0	(2)
15	Alaska	0	(5)
17	Taku	9	(26)
18	Stikine	7	(22)
19-23	Skeena	0	(12)
24	Alsek	28	(28)
50	Puget Sound	222	(85)
51	Juan de Fuca	0	(2)
52	Coastal Washington	776	(158)
53	Lower Columbia	273	(94)
54	Upper Columbia Spring	0	(4)
55	Upper Columbia Summer/Fall	3038	(269)
56	Snake Spring/Summer	18	(27)
57	Snake Fall	490	(167)
58	North & Central Oregon	1960	(227)
59	South Oregon Coastal	5	(29)
61	Klamath/Trinity	1	(8)
62	Middle Columbia Spring	0	(5)
63	Upper Willamette	34	(35)
64	Central Valley Fall	0	(6)
65	Central Valley Spring	0	(3)
66	Coastal California	0	(2)

FIGURES

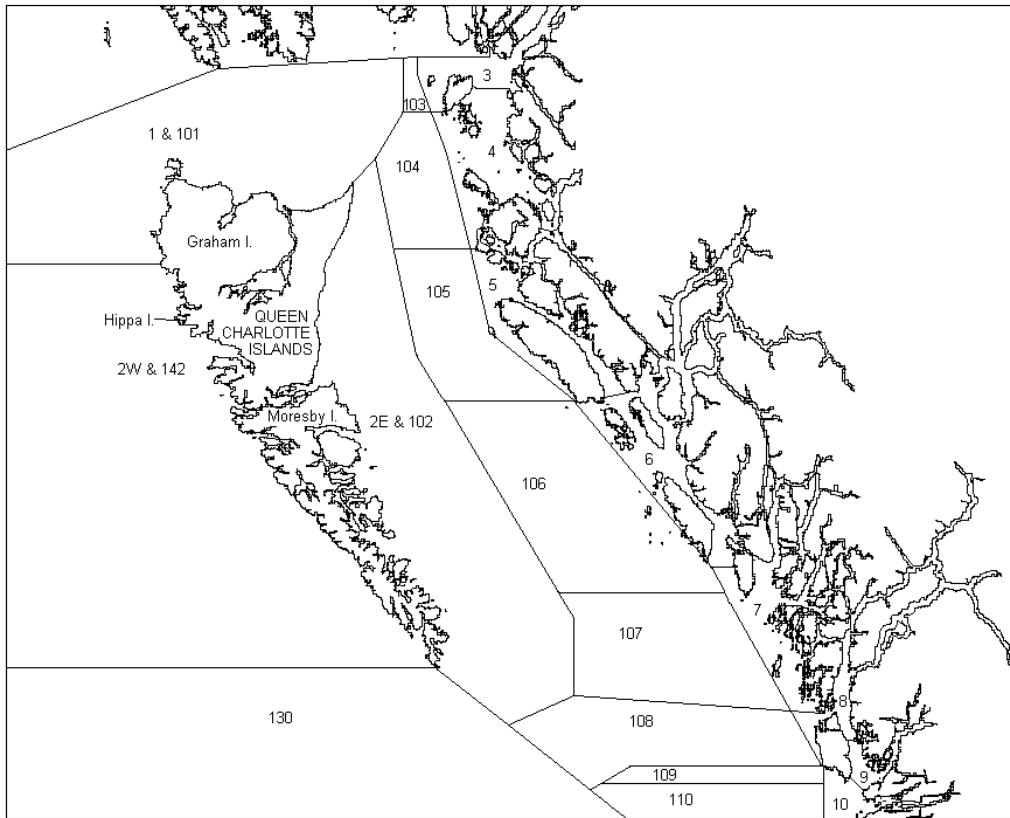


Figure 1. The North Coast of British Columbia showing Pacific Fishery Management Areas 1 to 10, 101 to 110, 130 and 142.

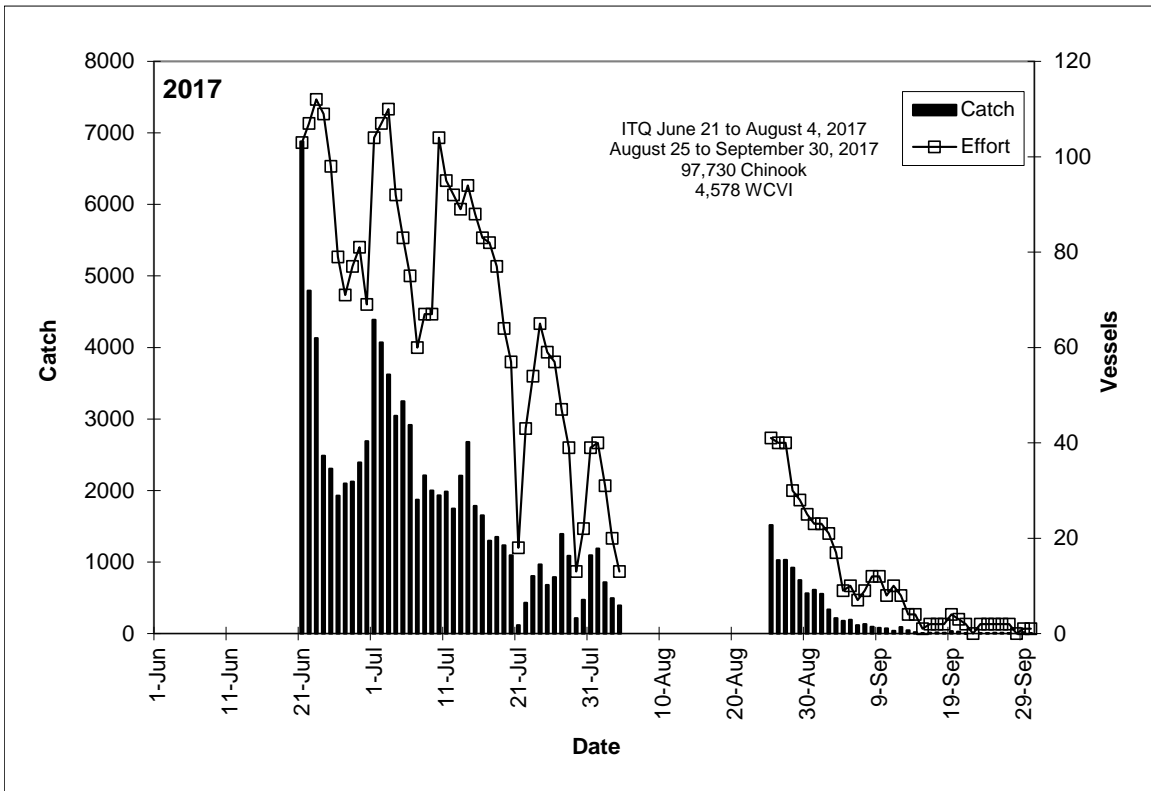


Figure 2. Chinook salmon daily catch and effort in the 2017 NBC Troll fishery.

APPENDICES

Appendix 1. Contract Statement of work for scale sampling of Chinook Salmon from the 2017 Northern British Columbia Troll fishery.

PERIOD: June 15 to September 30, 2017.

The contractor agrees to collect scale samples from Chinook salmon caught in the Northern British Columbia Troll fishery in conjunction with sampling for coded wire tags (CWTs) as described in this Statement of Work.

1. SCOPE

Genetic stock identification of Chinook salmon caught in the Northern British Columbia (NBC) troll fishery is used to establish the origin of fish encountered. The data generated by the program contribute to Canada's domestic management of the fishery. The information has also been used in support of Pacific Salmon Treaty (PST) negotiations. Genetic material and age information are available from scale samples.

The objectives for the genetic program are to provide estimates of the stock composition of Chinook caught in the NBC Troll fishery. These data are essential to the management of the fishery. The management objectives for the NBC Troll fishery are to catch as many of the Chinook salmon allowed within the PST allocation while remaining below a harvest rate of 3.2% of the WCVI Chinook return to Canadian waters. The preliminary estimate of the Chinook available to the NBC Troll fishery is 108,500 fish.

The NBC Troll Fishery is scheduled to open on or after June 21, 2017. The fishery may close near the end of July and reopen in September if the total allowable catch and the WCVI harvest rate objective have not been reached. The dates may change and the fishery will close once either of the targets has been met. A proposed licence condition will require all vessels to land their catch within 5 days of a closure.

The DFO Project Coordinator (Coordinator) for this SOW is Ivan Winther. The coordinator may be contacted by email at Ivan.Winther@dfo-mpo.gc.ca or by telephone at 250-627-3459. The shipping address is:

Ivan Winther
Fisheries & Oceans Canada
417-2nd Avenue West
Prince Rupert, BC
V8J 1G8

2. SAMPLE COLLECTION

The NBC Troll genetic sampling objectives are to collect scales and length data from a representative sample of the Chinook salmon catch, supply the samples with the associated capture information to the Coordinator, and provide standardized data to the DFO MRP Unit.

The collection of tissue samples preserved in ethanol has been discontinued as the preferred method for samples of genetic material. For Ice landings, genetic material is available from the *2017 NBC Troll Chinook Stock Composition* *NF-2017-VHP-14*

scales, however, it is subject to cross contamination from contact with other fish. In the revised scale sampling procedure the preferred location on each fish sampled will be wiped with a clean paper towel to avoid problems associated with cross contamination. For freezer landings, tissue samples are to be collected into batch vials containing water, transferred to Whatman paper and air-dried at the end of each day.

The sampling objective will be to collect a representative sample of 3000 Chinook salmon from the catch. This strategy is intensive in an attempt to measure the relatively small component of WCVI Chinook anticipated in the fishery. Sampling rate will be dictated by the rate of the fishery. It will be essential to sample heavily from the first landings of Chinook salmon and again once the fishery closes. A random / stratified approach is used to ensure a representative sample. Vessels are selected at random and every 5th fish or every 10th fish offloaded is sampled to reduce autocorrelation. Fewer than 50 fish are sampled from each vessel landing. Vessel operators are interviewed to determine areas of catch and number of Chinook being delivered. Samples of fresh landings are preferred as they provide the opportunity to sample for size, age and genetic information (as provided by length measurements and scales). The scale samples and data are matched with CWT recoveries when CWTs are encountered. Samples from fish frozen at sea are a second choice as they can only consist of genetic information available from tissue samples. Samples from frozen fish consist of batch collections of tissues into water and preserved on Whatman sheets. Scales and length information are not available from frozen fish.

2.1 The DFO Mark Recovery Unit shall:

- Supply the contractor with DFO Mark Recovery Sample Form (MRSF) Form DMP-MRP-FO-F-001 for CWT sampling and with the DFO Salmon Biological Sample Form (SBSF) Form DMP-MRP-FO-F-002 for DNA sampling.
- Supply the contractor with a set of waterproof labels to uniquely label each vial that is used for freezer vessel sampling.
- Provide a central repository for management and access to CWT and Salmon biological data collected for this project.

2.2 The Coordinator shall:

- Co-ordinate communication with the DFO Sclerochronology Lab (SCL) as the “DFO Submission Contact”.
- Supply single-type scale books (numbered 1-50) from the SCL to the contractor, for sampling fresh catch.
- Supply Whatman sheets to the contractor, for air-drying samples from frozen catch.
- Perform quality inspections to ensure samples meet current SCL or MGL identification and quality requirements and report deficiencies to the contractor.

2.3 The contractor shall:

- Communicate with the Coordinator or designate on appropriate sample times, locations and sample amounts.
- Examine scale books and provided by the Coordinator and report deficiencies to the coordinator verbally or by email.
- Collect scale samples from up to 3000 Chinook salmon following the instructions in section 3.

- Provide vials for sampling frozen fish.
- If requested, collect batched samples of tissue into water from frozen landings and transfer specimens onto Whatman paper.
- Ship or deliver the scale books, Whatman paper, and copies of completed SBSF's to the Coordinator within 2 days of sampling.
- Complete data entry of SBSFs and electronically submit data to the MRP Unit according to the following schedule:
 - June/July fishery by September 1, 2017
 - Late August/September fishery by November 1, 2017.
- Discuss modifications to procedures with the coordinator and incorporate modifications if necessary. If additional contract costs will be incurred, discuss modifications to procedures and cost estimate with Scientific Authority.
- Return all of the unused scale books and Whatman sheets to the coordinator at the end of the season.

3. SAMPLE INSTRUCTIONS

2017 North Coast Chinook Troll Commercial Fishery Samples

Samples of chinook tissue and scales will be collected from the Commercial Troll Fishery on the Queen Charlotte Islands. The sampling target is to collect 3000 scale samples from the Area F troll fishery in 2017.

- The sampling protocol is to collect scale samples with associated length measurements and capture data. The samples will be collected according to a stratified random procedure. Data from Chinook with CWTs will be matched to the scale sample data. Sample 50 or less Chinook from each vessel landing.
- Once a random vessel is selected to be sampled the scales and data are collected in a systematic procedure where every 5th or 10th Chinook from the load is sampled depending on the total number of fish delivered.
- The selection of Chinook for scale sampling shall be independent of whether a CWT is detected.

COLLECT SCALES AND CAPTURE DATA FROM EACH FISH ACCORDING TO THE FOLLOWING PROTOCOL:

- Use pencil.
- For each vessel, the MRSF (Form DMP-MRP-FO-F-001) will have been completed in full to record information from the vessel master interview and CWT sample data as part of the MRP Program requirements.
 - Complete the (SBSF Form DMP-MRP-FO-F-002) and link it to the MRSF data using the Master Sample # field according to the MRP Program requirements. In addition to MRP program requirements to link forms, copy the Fishing Location, Date Landed, First Day Fished, and Last Day Fished, from the MRSF onto the SBSF.
 - Complete the Sample Type to identify the sample protocols as “ 1-Random”.
- For scale sampling of ice vessel landings,
 - Record the Fork Length, Species, AD Fin Mark, Scale Book #, and Scale # for each fish sampled.

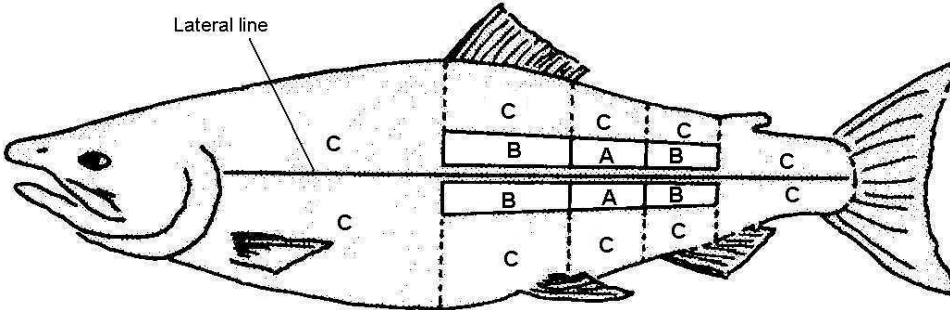
- Record the CWT Head Label Number for the scale samples of Chinook with a CWT.
- Each page will contain scales from 25 fish. Each vessel will have no more than 2 pages.
- Complete the Sample Status and provide a Sample Comment for incomplete or problem samples.
- Sample each fish in sequence according to the numbers on the scale books.
- **WIPE THE SLIME FROM THE PREFERRED SCALE LOCATION ON EACH FISH WITH A CLEAN SHEET OF PAPER TOWEL.**
- **DO NOT SAMPLE REGENERATE SCALES**
- Collect 5 scales from each fish onto standard 5-down format scale books as noted in the following illustration. Each book will contain scales from 10 fish.
- Ensure the numbers on the data sheets match the numbers on the scale books.
- For batch sampling of freezer vessel landings, Record the Species and vial number for the batch sample on the SBSF in the Sample Comments field.
 - Use a punch to collect a sample from each fish into a batch vial containing water.
 - **TAKE A SAMPLE FROM THE THINNEST PART OF THE CAUDAL FIN.** If the tissue is too thick it will not air dry effectively to stick to the Whatman sheet and can be knocked off.
 - Complete the Sample Status and provide a Sample Comment for incomplete or problem samples.
 - Ensure the vial number on the data sheet matches the number on the vial.
 - At end of day, transfer specimens from the batch vial to individual cells on the Whatman sheets. **SHORT-TERM** storage in water is OK but the sample will not stick to the Whatman paper if left in water overnight.
 - The sample should be placed in the centre of each of the squares printed on the paper. The paper will absorb all the water in the tissue causing it to adhere to the paper. Keep the sheet as dry as possible and avoid bending which might cause the samples to fall off.
 - Each Whatman page will contain samples from 100 fish. Each vessel will fill no more than ½ a Whatman sheet.
 - Record the date, species, fishing area and fishing gear on the Whatman Sheet.
 - Record the specimen numbers from the Whatman sheets onto the original SBSF. Each vessel will fill no more than 2 pages.
 - Ensure the specimen numbers on the Whatman sheets match the DNA specimen numbers on the data sheets.
 - Once the paper is completely dry it can be gently placed in the provided sleeves with the wax paper covering the samples. This can then be placed in padded or stay-flat envelopes for storage or shipping.
- Video: DNA sampling using Whatman paper: <http://www.youtube.com/watch?v=W9opRMYYVqU>

Copies of the completed SBSF data sheets must be shipped with the samples within two days of sampling.

SCALE SAMPLES:

- Record the scale book number on the data sheet.

- Scale samples must come from preferred locations on the fish as indicated by an "A" in the diagram below.
- Avoid collecting scales near scars, wounds or net marks. To avoid scars you may have to collect scales from the locations marked "B" in the diagram below.
- Do not collect scales from the lateral line or from the areas marked "C".



- **WIPE THE SLIME FROM THE PREFERRED LOCATION WITH A CLEAN SHEET OF PAPER TOWEL.**
- Using forceps collect a scale from the preferred location.
- **Please check for REGENERATE SCALES.** Check the scale to ensure the rings extend all the way to the center of the scale. If not, discard the scale and select another. Regenerated scales have a clear spot in the center of the scale which is missing the rings necessary to determine the age of the fish.
- Wipe off the scale and add it to the scale book on the appropriate numbered square, x, in the top row of the 10 X 5 scale book grid.
- Do not turn the scale over; leave the scale with the same side up as it is on the fish.
- Select 4 additional scales and add them to the adjacent squares below the first, bearing the same number series 1x, 2x, 3x, 4x.
- Keep the scale books dry.
- Once the book is full, fill out the information on the back of the page bearing the scales.
- **THE SCALES MUST MATCH THE NUMBERS ON THE DATA SHEETS.**

4. SAMPLE FORM

The following is an example of form DMP-MRP-FO-F-002 to be completed and provided with the sample collections.

Salmon Biological Sample Form

Sample #: 000000 Master Sample #

Fishery (*circle one*) **A B C D E F G H Other:** _____ Fishery Type Gear

Port _____ Sample Type Sample Status

Offload Site _____

VTP Vessel Name _____ VRN _____ Sample Site _____

FISHING LOCATION

PFMA	PFMA SUB	ZONE
1	-	-
2	-	-
3	-	-
4	-	-

Date Landed

d	d	m	m	y	y	y	y
---	---	---	---	---	---	---	---

First Day Fished

d	d	m	m	y	y	y	y
---	---	---	---	---	---	---	---

Last Day Fished

d	d	m	m	y	y	y	y
---	---	---	---	---	---	---	---

Date Sampled

d	d	m	m	y	y	y	y
---	---	---	---	---	---	---	---

	CWT Head Label #		Length (mm)		Species	AD Fin Mark	Sex	Scale Book #		Scale #	DNA Specimen #	DNA Tissue	Otolith Box #	Otolith Specimen		Comments
	Detection Method <input type="checkbox"/>	Electronic Equipment <input type="checkbox"/>	Length Code <input type="checkbox"/>					Book Type <input type="checkbox"/>	Format <input type="checkbox"/>					cell # <input type="radio"/>	strap # <input type="radio"/>	
1																
2																
3																
4																
5																
6																
7																
8																
9																
10																
11																
12																
13																
14																
15																
16																
17																
18																
19																
20																
21																
22																
23																
24																
25																

Sample Comments (for example: describe sample method, problems, etc.)

Sampling Agency _____

Lead Sampler _____

Recorded By _____

Stat Week Sampled _____

Page Number _____

DFO-201802

<p>FISHERY TYPE</p> <p>C – Commercial F – FN FSC N – FN Economic T – Test</p>	<p>GEAR</p> <p>7 – Hook & Line 10 – Gillnet 15 – Mixed Net 20 – Seine 31 – Freezer Troll 33 – Ice Troll</p>	<p>VTP</p> <p>1 – Vessel 2 – Truck 3 – Packer</p>
<p>SAMPLE TYPE</p> <p>1 – Random 2 – Total 3 – Stratified 4 – Random Stratified 5 – Selected 9 – Other</p>	<p>SAMPLE STATUS</p> <p>C – Complete I – Incomplete P – Problem <i>(Sample Comment Required for "I" and "P")</i></p>	
<p>DETECTION METHOD</p> <p>V – Visual (<i>Take AD-OFF/STUBBY</i>) E – Electronic (<i>Take BEEP</i>) B – Both (<i>Take AD-OFF/STUBBY that BEEP</i>)</p>		
<p>ELECTRONIC EQUIPMENT</p> <p>T – Tube W – Wand</p>		
<p>LENGTH CODE</p> <p>1 – Fork Length 7 – Post Orbital Fork 8 – Post Orbital Hypural</p>	<p>SPECIES</p> <p>08 – Pink 12 – Chum 15 – Coho 18 – Sockeye 24 – Chinook 28 – Steelhead</p>	<p>AD FIN MARK</p> <p>0 – AD-ON 1 – AD-OFF 2 – Stubby</p> <p>SEX</p> <p>F – Female M – Male</p>
<p>SCALE BOOK TYPE</p> <p>S – Single D – Double</p>	<p>SCALE FORMAT</p> <p>1 – 2 across 2 – 5 down 3 – 10 across 4 – 2 smears</p>	
<p>DNA TISSUE</p> <p>5 – adipose punch 6 – caudal or other fin punch 7 – scales 8 – operculum punch 9 – other</p>		

5. WHATMAN SHEET

The fields within each cell are duplicates of fields that are collected on the MRSF and do not need to be filled in. Follow procedures for sampling and shipping that are in this SOW.

Molecular Genetics Laboratory, Pacific Biological Station, Nanaimo
 For information contact: Kayla Mohan 250-756-7129 or John Candy 250-756-7224

Species: CN Vial Code:
 Area: F Samplers: JOT
 Year: Troll Catch Date: YYYY MM DD

	1	2	3	4	5	6	7	8	9	10
DAY AREA	DAY AREA	DAY AREA	DAY AREA	DAY AREA	DAY AREA	DAY AREA	DAY AREA	DAY AREA	DAY AREA	DAY AREA
Ad Cp Y / N	Ad Cp Y / N	Ad Cp Y / N	Ad Cp Y / N	Ad Cp Y / N	Ad Cp Y / N	Ad Cp Y / N	Ad Cp Y / N	Ad Cp Y / N	Ad Cp Y / N	Ad Cp Y / N
11	12	13	14	15	16	17	18	19	20	
DAY AREA	DAY AREA	DAY AREA	DAY AREA	DAY AREA	DAY AREA	DAY AREA	DAY AREA	DAY AREA	DAY AREA	DAY AREA
Ad Cp Y / N	Ad Cp Y / N	Ad Cp Y / N	Ad Cp Y / N	Ad Cp Y / N	Ad Cp Y / N	Ad Cp Y / N	Ad Cp Y / N	Ad Cp Y / N	Ad Cp Y / N	Ad Cp Y / N
21	22	23	24	25	26	27	28	29	30	
DAY AREA	DAY AREA	DAY AREA	DAY AREA	DAY AREA	DAY AREA	DAY AREA	DAY AREA	DAY AREA	DAY AREA	DAY AREA
Ad Cp Y / N	Ad Cp Y / N	Ad Cp Y / N	Ad Cp Y / N	Ad Cp Y / N	Ad Cp Y / N	Ad Cp Y / N	Ad Cp Y / N	Ad Cp Y / N	Ad Cp Y / N	Ad Cp Y / N
31	32	33	34	35	36	37	38	39	40	
DAY AREA	DAY AREA	DAY AREA	DAY AREA	DAY AREA	DAY AREA	DAY AREA	DAY AREA	DAY AREA	DAY AREA	DAY AREA
Ad Cp Y / N	Ad Cp Y / N	Ad Cp Y / N	Ad Cp Y / N	Ad Cp Y / N	Ad Cp Y / N	Ad Cp Y / N	Ad Cp Y / N	Ad Cp Y / N	Ad Cp Y / N	Ad Cp Y / N
41	42	43	44	45	46	47	48	49	50	
DAY AREA	DAY AREA	DAY AREA	DAY AREA	DAY AREA	DAY AREA	DAY AREA	DAY AREA	DAY AREA	DAY AREA	DAY AREA
Ad Cp Y / N	Ad Cp Y / N	Ad Cp Y / N	Ad Cp Y / N	Ad Cp Y / N	Ad Cp Y / N	Ad Cp Y / N	Ad Cp Y / N	Ad Cp Y / N	Ad Cp Y / N	Ad Cp Y / N
51	52	53	54	55	56	57	58	59	60	
DAY AREA	DAY AREA	DAY AREA	DAY AREA	DAY AREA	DAY AREA	DAY AREA	DAY AREA	DAY AREA	DAY AREA	DAY AREA
Ad Cp Y / N	Ad Cp Y / N	Ad Cp Y / N	Ad Cp Y / N	Ad Cp Y / N	Ad Cp Y / N	Ad Cp Y / N	Ad Cp Y / N	Ad Cp Y / N	Ad Cp Y / N	Ad Cp Y / N
61	62	63	64	65	66	67	68	69	70	
DAY AREA	DAY AREA	DAY AREA	DAY AREA	DAY AREA	DAY AREA	DAY AREA	DAY AREA	DAY AREA	DAY AREA	DAY AREA
Ad Cp Y / N	Ad Cp Y / N	Ad Cp Y / N	Ad Cp Y / N	Ad Cp Y / N	Ad Cp Y / N	Ad Cp Y / N	Ad Cp Y / N	Ad Cp Y / N	Ad Cp Y / N	Ad Cp Y / N
71	72	73	74	75	76	77	78	79	80	
DAY AREA	DAY AREA	DAY AREA	DAY AREA	DAY AREA	DAY AREA	DAY AREA	DAY AREA	DAY AREA	DAY AREA	DAY AREA
Ad Cp Y / N	Ad Cp Y / N	Ad Cp Y / N	Ad Cp Y / N	Ad Cp Y / N	Ad Cp Y / N	Ad Cp Y / N	Ad Cp Y / N	Ad Cp Y / N	Ad Cp Y / N	Ad Cp Y / N
81	82	83	84	85	86	87	88	89	90	
DAY AREA	DAY AREA	DAY AREA	DAY AREA	DAY AREA	DAY AREA	DAY AREA	DAY AREA	DAY AREA	DAY AREA	DAY AREA
Ad Cp Y / N	Ad Cp Y / N	Ad Cp Y / N	Ad Cp Y / N	Ad Cp Y / N	Ad Cp Y / N	Ad Cp Y / N	Ad Cp Y / N	Ad Cp Y / N	Ad Cp Y / N	Ad Cp Y / N
91	92	93	94	95	96	97	98	99	100	
DAY AREA	DAY AREA	DAY AREA	DAY AREA	DAY AREA	DAY AREA	DAY AREA	DAY AREA	DAY AREA	DAY AREA	DAY AREA
Ad Cp Y / N	Ad Cp Y / N	Ad Cp Y / N	Ad Cp Y / N	Ad Cp Y / N	Ad Cp Y / N	Ad Cp Y / N	Ad Cp Y / N	Ad Cp Y / N	Ad Cp Y / N	Ad Cp Y / N

Place a punch of thin absorbent or tea fin tissue in center of grid. To reduce cross-contamination, please wipe down the tissue site on the fish prior to sampling. Please ensure sampling tools have been rinsed between fish.

Appendix 2. Baseline samples used in the mixture analyses.

#	Region	Population	N				
1	UPFR	Bowron	250	6	LWTH	Louis	618
1	UPFR	Dome	382	6	LWTH	Nicola	433
1	UPFR	Fontoniko	63	6	LWTH	Spius	137
1	UPFR	Goat	76	6	LWTH	U_Coldwat_SP	221
1	UPFR	Holliday_Cr	29	6	LWTH	U_Spius_SP	175
1	UPFR	Holmes	219	7	ECVI	Big_Qualicum	365
1	UPFR	Horsey	47	7	ECVI	Chemainus	261
1	UPFR	Indianpoint	47	7	ECVI	Cowichan	680
1	UPFR	James	58	7	ECVI	L_Qualicum	305
1	UPFR	Kenneth_Cr	98	7	ECVI	Nanaimo_F	523
1	UPFR	McGregor	125	7	ECVI	Nanaimo_SP	95
1	UPFR	Morkill	208	7	ECVI	Nanaimo_SU	459
1	UPFR	Nevin_Cr	50	7	ECVI	NanaimoSu_BackX	135
1	UPFR	Ptarmigan	32	7	ECVI	Nimpkish	316
1	UPFR	Salmon@PG	263	7	ECVI	Puntledge_BackX	1025
1	UPFR	Slim_C	240	7	ECVI	Puntledge_F	652
1	UPFR	Swift	448	7	ECVI	Puntledge_Su	1120
1	UPFR	Tete_Jaune	475	7	ECVI	Quatse	30
1	UPFR	Torpy	174	7	ECVI	Quinsam	503
1	UPFR	Walker	45	7	ECVI	Woss_Lake	31
1	UPFR	Willow_R	117	8	WCVI	Burman	315
2	MUFR	Baezaeko	82	8	WCVI	Colonial_Cay	58
2	MUFR	Baker_Cr	31	8	WCVI	Conuma	455
2	MUFR	Bridge	424	8	WCVI	Gold_R	227
2	MUFR	Chilako	45	8	WCVI	Kaouk_R	196
2	MUFR	Chilko	425	8	WCVI	Kennedy	383
2	MUFR	Cottonwood	176	8	WCVI	Marble@NVI	512
2	MUFR	Elkin_R	248	8	WCVI	Megin_R	90
2	MUFR	Endako	207	8	WCVI	Moyeha_R	57
2	MUFR	Horsefly	80	8	WCVI	Nahmint	411
2	MUFR	Kuzkwa_Cr	93	8	WCVI	Nitinat	346
2	MUFR	L_Cariboo	104	8	WCVI	Robertson	388
2	MUFR	L_Chilcoti	236	8	WCVI	San_Juan	202
2	MUFR	Nazko	194	8	WCVI	Sarita	429
2	MUFR	Nechako	562	8	WCVI	Sooke	58
2	MUFR	Portage_C	286	8	WCVI	Stamp	299
2	MUFR	Quesnel	562	8	WCVI	Stamp	299
2	MUFR	Stuart	545	8	WCVI	Tahsis	355
2	MUFR	Taseko	205	8	WCVI	Thornton	522
2	MUFR	U_Cariboo	171	8	WCVI	Tlupana	66
2	MUFR	U_Chilcotin	276	8	WCVI	Toquart	87
2	MUFR	Westroad	104	8	WCVI	Tranquil	395
3	LWFR-F	Chilliwac@Stav	381	9	SOMN	Zeballos	199
3	LWFR-F	Chilliwack_F	696	9	SOMN	Bute	66
3	LWFR-F	Harrison	686	9	SOMN	Capilano	126
4	NOTH	Barriere	55	9	SOMN	Cheakamus	50
4	NOTH	Blue	84	9	SOMN	Cheakamus_F	114
4	NOTH	Clearwater	281	9	SOMN	Cheakamus_Su	40
4	NOTH	Finn	211	9	SOMN	Devereux	325
4	NOTH	Lemieux_Cr	153	9	SOMN	Homathko	51
4	NOTH	N_Thom@Main	116	9	SOMN	Klinaklini	472
4	NOTH	Raft_R	457	9	SOMN	Mamquam	35
5	SOTH	Bessette	164	9	SOMN	Phillips	641
5	SOTH	Duteau_Cr	73	9	SOMN	Squamish_R	161
5	SOTH	Eagle_R	331	10	NOMN	Ashlulm	66
5	SOTH	L_Adams	340	10	NOMN	Atnarko	275
5	SOTH	L_Shus@U_Adams	46	10	NOMN	Chuckwalla	315
5	SOTH	L_Shuswap	389	10	NOMN	Dean	219
5	SOTH	L_Thompson	229	10	NOMN	Dean@Main	25
5	SOTH	Little_R	254	10	NOMN	Docee	126
5	SOTH	M_Shuswap	375	10	NOMN	Hirsch	474
5	SOTH	Salmon@SA	215	10	NOMN	Kateen	244
5	SOTH	Seymour@Thomp	44	10	NOMN	Kilbella	196
5	SOTH	South_Thom	266	10	NOMN	Kildala	441
6	LWTH	Bonaparte	344	10	NOMN	Kitimat	483
6	LWTH	Coldwater	274	10	NOMN	Kitlope	201
#	Region	Population	N	#	Region	Population	N
6	LWTH	Deadman	492	10	NOMN	Kwinamass	362
				10	NOMN	LowAtnarko	50
				10	NOMN	Marble@CC	41
				10	NOMN	Neechanze	57

10	NOMN	Nusatsum	103	23	Skeena Lower	Fiddler_Cr	113
10	NOMN	Saloompt	138	23	Skeena Lower	Gitnadoix	245
10	NOMN	Takia	63	23	Skeena Lower	Kasiks_R	63
10	NOMN	U_Atnarko	200	23	Skeena Lower	Khyex_R	37
10	NOMN	U_Dean	203	23	Skeena Lower	Kitsumkalum_R	810
10	NOMN	Wannock_R	506	23	Skeena Lower	Thomas_Cr	117
11	NASS	Cranberry	175	23	Skeena Lower	Zymogotitz_R	120
11	NASS	Damdochax	273	24	Alsek	Blanchard	381
11	NASS	Ishkheenickh	199	24	Alsek	Goat_Cr	134
11	NASS	Kincolith	286	24	Alsek	Klukshu	433
11	NASS	Kiteen	59	24	Alsek	Kudwat_Cr	70
11	NASS	Kwinageese	266	24	Alsek	Takhanne	218
11	NASS	Meziadin	194	24	Alsek	Tatshenshi	24
11	NASS	Owegee	235	50	Puget Sound	Green@Kendal_F	50
11	NASS	Seaskinnish	99	50	Puget Sound	Green_F@Soos	100
11	NASS	Snowbank	51	50	Puget Sound	Nooksack_SP@Ke	200
11	NASS	Teigen	30	50	Puget Sound	Serpentine	46
11	NASS	Tseax	244	50	Puget Sound	Skagit_Su	310
12	LWFR-Sp	Big_Silver	210	50	Puget Sound	Skykomish_Su	114
12	LWFR-Sp	Birkenhead	347	50	Puget Sound	Snohomish_R	306
12	LWFR-Sp	BlueCr_UpPitt	50	50	Puget Sound	Soos_Cr_H	183
12	LWFR-Sp	Sloquet_Cr	35	50	Puget Sound	StillaguamishS	87
12	LWFR-Sp	Upper_Pitt	235	50	Puget Sound	White_F	252
13	LWFR-Su	Maria_Slough	366	51	Juan de Fuca	Elwha_F	99
13	LWFR-Su	Nahatlatch_R	26	52	Coastal Wash	Hoh_River_SP_S	59
14	QCI	Yakoun	211	52	Coastal Wash	Queets	138
15	Alaska	Big_Boulder_C	144	52	Coastal Wash	Quinault_F	100
15	Alaska	Chickamin	259	52	Coastal Wash	Solduc_F	98
15	Alaska	King_Salmon	266	52	Coastal Wash	Willapa_Cr	261
15	Alaska	Situk	132	53	Low Col	Abernathy_F	100
15	Alaska	Tahini	142	53	Low Col	Coweeman	195
15	Alaska	Unuk	336	53	Low Col	Cowlitz_H_Sp	138
17	Taku	Dudidontu	352	54	Up Col-Sp	Chewuch_SP	100
17	Taku	Hackett_r	233	54	Up Col-Sp	Chiwawa_SP	100
17	Taku	Kowatua	379	54	Up Col-Sp	Entiat_SP	142
17	Taku	Little_Tatsam	698	54	Up Col-Sp	Twisp_SP	227
17	Taku	Nahlin	303	55	Up Col-Su/F	Deschutes-F	230
17	Taku	Nakina	480	55	Up Col-Su/F	Hanford_Reach	617
17	Taku	Tatsamenie	38	55	Up Col-Su/F	Okanagan	132
17	Taku	Tseta	327	55	Up Col-Su/F	Osoyoos_Resid	35
17	Taku	Yeth_Cr	53	55	Up Col-Su/F	Silmilkameen_S	370
18	Stikine	Andrew_Cr	144	55	Up Col-Su/F	Wenatchee_Su	235
18	Stikine	Christina	240	56	Snake-Sp/Su	Frenchman-SP	61
18	Stikine	Craig	114	56	Snake-Sp/Su	Imnaha	239
18	Stikine	Johnny_Tashoot	99	56	Snake-Sp/Su	Johnson_Cr	240
18	Stikine	Little_Tahltan	745	56	Snake-Sp/Su	Marsh_Cr	220
18	Stikine	Shakes_Cr	225	56	Snake-Sp/Su	McCall	32
18	Stikine	Tahltan_R	212	56	Snake-Sp/Su	McCall_Hat	41
18	Stikine	Verrett	854	56	Snake-Sp/Su	Minam_Cr	144
19	Skeena Upper	Bear	270	56	Snake-Sp/Su	Rapid_Sp	363
19	Skeena Upper	Kluatantan	38	56	Snake-Sp/Su	Salmon_E_Fork	53
19	Skeena Upper	Kluayaz_Cr	165	56	Snake-Sp/Su	Secech	277
19	Skeena Upper	Kuldo_C	171	56	Snake-Sp/Su	Snake_S	36
19	Skeena Upper	Otsi_Cr	276	56	Snake-Sp/Su	Tucannon_SP	274
19	Skeena Upper	Sicintine_R	319	56	Snake-Sp/Su	Up_Salmon-SP	165
19	Skeena Upper	Slamgeesh	129	56	Snake-Sp/Su	Upper_Valley	77
19	Skeena Upper	Squingula_R	271	56	Snake-Sp/Su	Valley_Cr	43
19	Skeena Upper	Sustut	509	56	Snake-Sp/Su	Wenaha	89
20	Skeena Babine	Babine	198	57	Snake-F	Lyon's_Ferry_F	370
21	Skeena Bulkley	Bulkley_Early	567	58	North & Central Oregon	Cle_Elm_Hatch	95
21	Skeena Bulkley	Morice_R	243				
21	Skeena Bulkley	Suskwa	111	58	North & Central Oregon	Elk	206
22	Skeena Mid	Kispiox	197				
22	Skeena Mid	Kitseguecla_R	260	58	North & Central Oregon	Euchre_Cr	57
22	Skeena Mid	Kitwanga	284				
22	Skeena Mid	Nangeese_R	32	58	North & Central Oregon	Nehalem	327
22	Skeena Mid	Shegunia_R	132				
22	Skeena Mid	Sweetin	245				
#	Region	Population	N	#	Region	Population	N
23	Skeena Lower	Cedar_Early	116	58	North & Central Oregon	Siuslaw	258
23	Skeena Lower	Ecstall	367				
23	Skeena Lower	Exchamsiks	116	58	North & Central Oregon	Trask_hat_F	236
23	Skeena Lower	Exstew_R	140				

58	North & Central Oregon	Trask_hat_SP	48
58	North & Central Oregon	Umpqua_Smith	229
59	South Oregon coastal	Cole	188
59	South Oregon coastal	Hunter_Cr	96
59	South Oregon coastal	Lobster_Cr	49
59	South Oregon coastal	Nestucca_F	153
59	South Oregon coastal	Pistol	98
59	South Oregon coastal	Umpqua_Sp	136
59	South Oregon coastal	Winchuk	80
61	Klamath/Trinity	Blue_Cr	94
61	Klamath/Trinity	Salmon_Cal	28
61	Klamath/Trinity	Trinity_F	244
61	Klamath/Trinity	Trinity_SP	100
62	Mid Col-Sp	Granite	93
62	Mid Col-Sp	John_Day_main	228
62	Mid Col-Sp	John_Day_Mid	40
62	Mid Col-Sp	John_Day_N	40
62	Mid Col-Sp	Naches_Sp	109
62	Mid Col-Sp	Spring_Cr_H	137
63	Up Willamette	Clackamas_N	79
63	Up Willamette	North_Santiam	236
63	Up Willamette	Sandy	208
64	Cent Val-F	American	69
64	Cent Val-F	Battle_Cr	183
64	Cent Val-F	Butte_F	49
64	Cent Val-F	Feather_F	272
64	Cent Val-F	Merced	200
64	Cent Val-F	Mokelumne	95
64	Cent Val-F	Sacr_F	129
64	Cent Val-F	Sacr_LF	211
64	Cent Val-F	Stanislaus	101
64	Cent Val-F	Toulumne	34
64	Cent Val-F	Yuba	50
65	Cent Val-Sp	Butte_Sp	186
65	Cent Val-Sp	Feather_Sp	226
65	Cent Val-Sp	Yuba_Sp	32
66	Coastal California	Eel_F	279

