# Statistical basis for use of genetic information for cohort analysis. 

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Using DNA to develop stock composition estimates
populations from
SE Alaska to Cal "inseason"
turnaround 24-48 hrs.

Mixture
Estimate

Application of mixed stock genetic data now
Adaptive fisheries management


Can we use this data for more than fisheries management purposes?

We require for cohort analysis:
-Accurate estimates to stock-of-origin.
-Accurate estimates of age
Pre-season planning

CWT "like" data from DNA sampling


## Accuracy and Precision

-Estimation Error
-What is the stock composition accuracy in the sample?
(accuracy ~ baseline build)

- What is the trade-off between sample size and precision in the model?
-Testing accuracy using known mixtures.


## Model Accuracy and Precision - baselines

Standard baseline

- Not all mixture populations may be present in the baseline. We rely on populations in the baseline to stand proxy for those populations not in the baseline. We rely on "regional structure" to make accurate assignments to a region. Estimate stock compositions to regions only. Reasonably accurate estimate to region (fisheries management applications).

Optimized baseline

- Three criteria met: 1) All mixture populations are represented in the baseline; 2) Enough genetic markers used to provide sufficient discrimination between populations; 3) baseline populations have large enough sample size.
- Should be able to accurately estimate to population.

Model Accuracy and Precision - baselines
Optimized baseline - Sufficient number of markers to allow separation between populations.

Some considerations:
-not all markers are equal (highly polymorphic markers provide more information)
-adding markers cost more work and more $\$$.
-certain markers can be run together (multiplex) saves \$.


## Model Accuracy and Precision - baselines

## Optimized baseline - Baseline populations have sufficient sample size.



## Model Accuracy and Precision - mixtures

-Relationship between mixture sample size and SD
-Simulation for three populations at 100\%


Known Sample - CTC Multi-Agency Sampling plan

| Region | Numbers of <br> populations | Number of <br> individuals | Source <br> laboratory |
| :--- | :---: | :---: | :--- |
| Kamchatka to SE | 10 | 100 | ADFG/ABL |
| Alaska | 10 | 100 | CDFO |
| British Columbia | 5 | 50 | WDFW |
| Puget Sound |  |  | WDFW/ |
| NWFSC/CRITFC/ |  |  |  |
| Columbia River | 15 | 150 | UI/IDFG |
| Washington/Uregon | 5 | 50 | WDFW/OSU |
| Coast <br> California | 5 | 50 | SWFSC |

## Example 1 - Model Accuracy (standard and optimized baseline)



Example 1 - Individual assignments from "optimized" portion of baseline - Multi-Agency mixture sample .

| Region | Population | Correct to <br> Population | Correct to <br> Region | Total |  |
| :--- | :--- | :---: | :---: | :---: | :---: |
| SEAlaska | King Salmon | 10 | 10 | 10 |  |
| Skeena | Sustut | 10 | 10 | 10 |  |
| Nass | Tseax | 10 | 10 | 10 |  |
| CCST BC | Klinaklini | 8 | 8 | 10 |  |
| For individual |  |  |  |  |  |
| assignments take |  |  |  |  |  |
| the highest bayes |  |  |  |  |  |
| probability |  |  |  |  |  |
| associated with a |  |  |  |  |  |
| population for |  |  |  |  |  |
| each fish. |  |  |  |  |  |
| WCVI | Dean | 8 | 8 | 10 |  |
| WCVI | Sarita | 9 | 10 | 10 |  |
| ECVI | Tahsis | 10 | 10 | 10 |  |
| LWFR | Nanaimo | 10 | 10 | 10 |  |
| SOTH | Chilliwack | 7 | 7 | 9 |  |
| UPFR | Bessette | 10 | 10 | 10 |  |
|  | Holmes | 9 | 9 | 10 |  |
| Total |  |  |  |  |  |

Example 1 - Individual assignments from "standard" portion of baseline where baseline contains mixture population - MultiAgency mixture sample .

| Region | Population | Correct to <br> Population | Correct to <br> Region | Total |
| :--- | :--- | :---: | :---: | :---: |
| PugetSnd | Skagit | 5 | 5 | 6 |
| PugetSnd | Green | 9 | 10 | 10 |
| PugetSnd | White | 9 | 10 | 10 |
| Coast Wash | Hoh | 8 | 10 | 10 |
| Coast Wash | Quinault | 8 | 9 | 10 |
| Low Col/Will | Abernathy | 9 | 10 | 10 |
| Snake-F | Deschutes_F | 6 | 8 | 10 |
| UpCol-Su/F | Handford Reach | 5 | 9 | 10 |
| Snake-Sp/S | South_F_Salmon | 10 | 10 | 10 |
| Snake-Sp/S | Mid_F_Salmon | 8 | 10 | 10 |
| Snake-Sp/S | Rapid | 10 | 10 | 10 |
| Snake-Sp/S | Imnaha | 1 | 10 | 10 |
| Snake-F | Lyons Ferry | 2 | 2 | 9 |
| MidCol-Sp | Granite | 8 | 8 | 10 |
| Low Col/Will | Mackenzie | 0 | 9 | 10 |
| CentVal_F | Sacr_F | 2 | 4 | 10 |
| Cen_Val_Sp | Bute | 7 | 9 | 10 |
| Total | 107 | 143 | 165 |  |
| Percent |  |  |  |  |

## Example 2 - Model accuracy - standard baseline -known CWT

 mixture sample

## Example 3-Optimized baseline

Fraser chinook migration timing at Albion.

Baseline Fraser 56 populations
Open boxes $=4,822$ DNA individual assignments (2000-2001)

Dark boxes = 516 CWT (1987-2003)
Parken et al. (in prep)

Most of the comparisons between CWT and DNA show no significant difference between median migration timing, where they do we can usually identify inadequacies in the baseline.


## DNA implementation considerations - sample size

Use sampling theory to determine precision levels for given sample sizes.


DNA implementation considerations - random vs. non-random sampling.
"Boat Effect" Sockeye example
Three seine boats, all fishing Area20 on August 12, 2003
Chi-squared value $=3.33$ (Critical value $p<0.05=9.49$ )


DNA implementation considerations - information systems.
User interface


## DNA cost.

Depends on numbers of samples analyzed:
-Levels of stratification required (area, gear, age, time, marked/unmarked)
-Sampling precision (sampling theory)
-Coordination with Fisheries Management sampling?
-Only interested marked fish and indicator stocks?

## DNA data available from BC fisheries - a starting point?

Some years there are more chinook DNA samples analyzed than CWT recovered, most years there is around $1 / 3-1 / 2$ as many DNA samples.

Not all this data has corresponding age


## Summary

## Benefits/Opportunities using DNA are:

-Collecting information on all populations in the catch, not just the tagged component.
-Regional groups correspond to genetic ESUIMU designations or rolled up into PSC regional groups.

- Optimized baseline will provide accuracy of $\sim 90 \%$ for individual populations (individual assignments) (consistent with what we see in sockeye).
-Room for improvement if the baseline can be reduced to indicator populations only.
-Ease of DNA sampling should allow for better random sampling of catch (sampling does not devalue fish).
-Get a handle on sub-legals, non-retention fish by non-lethal DNA sampling.
-Reconstruct historical fisheries using DNA from scales.


## Disadvantages using DNA are:

-Cannot separate hatchery and wild from the same population
-Must deal with an estimate of age and stock-of-origin if we want provide a CWT equivalent.

